

A Concept of a Probable Autoinjector for Bio-threat Agents

R.V. Geetha¹, Anitha Roy¹, S. Senthilkumar¹, A.S.B. Bhaskar[#], and R. Vijayaraghavan^{*1}

¹Saveetha University, Chennai – 602 105, India

[#]Defence Research and Development Establishment, Gwalior – 474 002, India

^{*}E-mail: jai_vijay@hotmail.com

ABSTRACT

Chemical and biological weapons can be used during conflicts and by terrorists to injure or kill humans and animals. Chemical weapons contain toxic chemicals and biological weapons contain pathogenic organisms. With proper protective equipments and training, the lethal effects of chemical and biological warfare agents can be minimised. First aid kit is available that contain detection, decontamination and medical protection for chemical warfare agents including autoinjectors, for rapid administration and faster absorption of drugs. The autoinjectors are safe and available for life saving drugs like atropine sulphate and pralidoxime chloride (nerve agent poisoning), epinephrine (anaphylaxis), diazepam (seizures) and sumatriptan (migraine). For bio-threat agents doxycycline alone is available as a broad spectrum antibiotic in the first aid kit. Majority of the bacterial agents are gram negative and hence amikacin drug cartridge was developed for the existing autoinjector. The advantage of amikacin is its safety, stability, can be given prophylactically and supplemented with other antibiotics when medical attention arrives. The usability and tolerability of amikacin administered repeatedly through autoinjector was studied using various haematological, biochemical and oxidative stress parameters in animal models. The results are promising and as there is no antibiotic autoinjector available, amikacin autoinjector can be considered for bio-threat agents.

Keywords : Autoinjector, bio-threat agents, amikacin, oxidative stress

1. INTRODUCTION

The use of chemical and biological warfare agents on humans, animals and plants to cause harm or death is a concept followed by several State Parties and by terrorist organisations¹. The chemical weapons can cause death and injury through its chemical action on the biological system². The biological weapons are bacterial and viral agents that are pathogenic to humans and animals, and toxins derived by biological sources³. The chemical agents act very fast and its detection and protection has to be carried out very quickly. Defence Research and Development Organisation has developed a first aid kit that contains detection, decontamination and medical protection⁴. The nerve agents like tabun, sarin, soman and VX act rapidly and hence for medical protection the antidotes are provided as autoinjectors in the first aid kit, for rapid administration and faster absorption of drugs.

The autoinjectors are very safe. When given intramuscularly the autoinjector can deliver the drugs faster with rapid absorption compared to manual injection due to the spray effect of drug delivery in autoinjector. In the autoinjector, the needle is not visible outside and hence do not cause any fear. The injection will be painless, easy to administer and well suited for mass casualty management. When healthy volunteers were administered atropine and pralidoxime chloride through autoinjectors, the drug absorption was faster by the autoinjector⁵. Other than atropine sulphate and pralidoxime

chloride (nerve agent antidotes), autoinjectors are available for few life saving situations viz., epinephrine for anaphylaxis, diazepam for status epilepticus, sumatriptan for migraine and midazolam for convulsions⁶.

For bio-threat agents in the first aid kit, doxycycline alone is available as a broad spectrum antibiotic for oral administration. During training and operation in the military service, low intensity conflicts, road accidents and also in natural disaster situations, serious injury may occur for which an antibiotic may be required. An autoinjector will be very useful in such situations. In addition, the antibacterial autoinjector can also be used as an antidote for bio-threat agents. A concept of development of a probable autoinjector for an antibacterial drug and an analgesic drug was earlier proposed^{6,7}. Previously, amikacin drug cartridge was developed for the existing reusable autoinjector and tested in animal models for its tolerance and usability using various haematological and biochemical parameters⁸. All parameters viz., liver function test, kidney function test, serum enzymes and general biochemical variables did not show any significant change⁹. Since the autoinjector delivers the drug with force, in the present study amikacin was delivered intraperitoneally in rats by autoinjector and compared with manual injection for various oxidative stress parameters. Since, amikacin will be effective against most of the gram negative organisms and few gram positive organisms, an *in vitro* study was also carried out using combination of antibiotics for its effectiveness for future development¹⁰.

2. MATERIAL AND METHODS

2.1. Animals

Female Wistar rats weighing 120 g - 180 g from Biomedical Research Unit and Laboratory Animal Centre, Saveetha University were used. The animals were housed in polypropylene cages, three per cage with dust free paddy husk as the bedding material. They were provided with laboratory animal feed (VRK Nutritional, India) and filtered water *ad libitum*. The animals were maintained at a temperature of 25 ± 2 °C and natural dark/light cycle. The study was approved by Institutional Animal Ethical Committee of Saveetha University (SU/ BRULAC/RD/010/2014).

2.2. Drug Cartridge and Dilution

Amikacin drug cartridge developed by Saveetha University with the collaboration of Defence Research and Development Organisation and M/s Neon Laboratories (Mumbai), was used for the study (as shown in Fig. 1). The drug cartridge was developed with human dosage of 250 mg/mL (2.35 mL/vial). For the animal administration, the human dose cartridge was diluted 1:4, to make the concentration to 63 mg/mL. For this, 1.75 mL of the drug solution was withdrawn from the cartridge in sterile condition using a laminar flow and 1.75 mL of sterile saline was injected back. The autoinjector capable of delivering full dose with full needle out or partial dose with partial needle out was used⁹. The drug cartridge was loaded in the autoinjector with the plastic clip restrictor for partial dose delivery (1.2 mL against 2.0 mL) and partial needle out (15 mm against 25 mm), since it was an animal study through intraperitoneal route⁹. The refilled amikacin cartridges were subjected to a variety of quality control parameters viz., sterility test, firing efficacy test, stability test, low pressure test and vibration test.

2.3. Experimental Groups

Thirty five female rats were randomly allocated to the following five groups (7 rats per group).

- Group 1 : Control
- Group 2 : Amikacin, 63 mg/mL (1.2 mL, i.p.) for 3 days by autoinjector,
- Group 3 : Amikacin, 63 mg/mL (1.2 mL, i.p.) for 3 days by manual injection.
- Group 4 : Amikacin, 63 mg/mL (1.2 mL, i.p.) for 7 days by autoinjector
- Group 5 : Amikacin, 63 mg/mL (1.2 mL, i.p.) for 7 days by manual injection.

The rats were held on the surgical table with its back on the table gently and firmly. The autoinjector was unlocked and positioned gently on the lower abdomen vertically. The trigger button was pressed and held on to the abdomen for 10 seconds. The drug cartridge was weighed before and after injection to estimate the quantity of the drug delivered. For the manual injection, the drug solution that was removed from the cartridge, was diluted suitably with sterile normal saline to get a concentration of 63 mg/mL and from that 1.2 mL was injected intraperitoneally (i.p.) to each rat.

2.4. Sample Collection

The animals were weighed daily. General behaviour, food and water intake were recorded. Twenty four hours after the last dose (4th or 8th day), the animals were anaesthetised with isoflurane and blood was collected from the orbital sinus in two separate tubes, one with anticoagulant (K₃EDTA) and the other without anticoagulant. After the collection of the blood, the animals were sacrificed with a over dose of anaesthesia. Vital organs like heart, lungs, spleen, kidney and liver were excised, blotted free of blood, weighed and preserved in formalin solution.

2.5. Oxidative Stress Parameters in Blood

Catalase (CAT), glutathione peroxidase (GPX), superoxide dismutase (SOD) and glutathione reductase (GR), were estimated using the haemolysate. From the tubes without anticoagulant, serum was separated and was used for the estimation of malondialdehyde (MDA) and reduced glutathione (GSH). Roche's diagnostic kits (Chennai, India) were used and the assays were carried out as per the manufacturer's instruction (expressed as units), in a spectrophotometer (Shimadzu, Japan).

2.6. *In vitro* Test for Combination of Antibacterial Agents

Disc diffusion test was performed according to the NCCLS procedure¹¹. The lawn culture of the test organisms were compared to 0.5 Mac Farland standard made on Mueller Hinton agar (HiMedia, India)¹². The antibacterial activity of the antibiotics were tested alone and also in combination. The plates were incubated at 37 °C for 24 h and the zone of inhibition was measured. Two strains were used for the *in vitro* sensitivity test, (i) *Pseudomonas aeruginosa* as gram negative and (ii) *Staphylococcus aureus* as gram positive. Amikacin alone and in combination with cefazolin and vancomycin were tested. The tests were done in triplicates.



Figure 1. Amikacin drug cartridges for reusable autoinjector, (a) fresh cartridges, (b) partially ejected drug cartridges, and (c) fully ejected drug cartridges.

2.7. Statistical Analysis

All the parameters were analysed using one way analysis of variance (ANOVA) and compared with control using Dunnett's test. A probability of 0.05 and less was taken as statistically significant. The analysis and plotting of graphs were carried out using SigmaPlot 12 (SysStat Software Inc., USA).

3. RESULTS

All the animals survived in all the groups. Bleeding or abscess was not observed in any of the groups. The food and water intake were also apparently similar in all the groups. The approximate dose of amikacin administered was 500 mg/kg and the effective needle length was 15.5 mm. The refilled cartridges complied with all the quality control parameters.

There was no significant change in blood GSH and MDA levels in 3 d and 7 d groups of autoinjector and manual injection when compared to control group as shown in Fig. 2. Similarly, no significant change was observed in blood SOD, CAT, GPX and GR levels in 3 d and 7 d groups of autoinjector and manual injection when compared to control group as shown in Figs. 3 and 4.

The results of *in vitro* test for the antibiotics and their combination against gram negative and gram positive organisms are given in Table 1. In the case of gram negative organism, amikacin alone or its combination with cefazolin or vancomycin showed no statistically significant difference, suggesting that amikacin alone is adequate. The combination did not show any antagonist effect against the gram negative organism. But, in the case of gram positive organism, combination with cefazolin or vancomycin showed statistically significant effect.

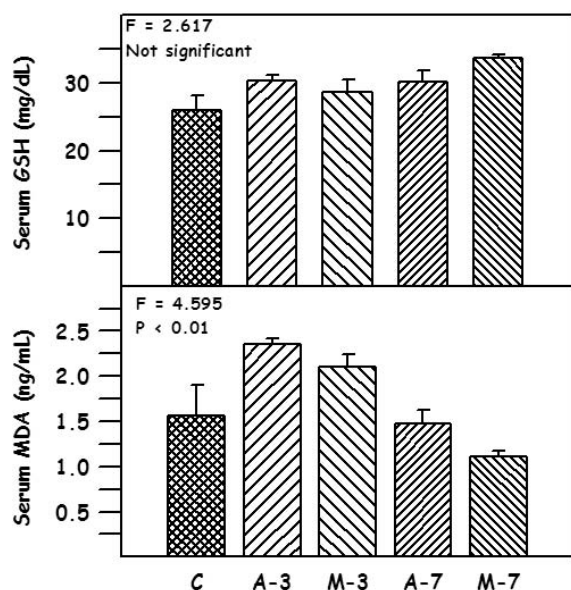


Figure 2. Effect of amikacin administered through autoinjector and manual injection by i.p. in rat on reduced glutathione (GSH) and malondialdehyde (MDA), C = Control Group; A-3 = autoinjector 3 doses; M-3 = manual injection 3 doses; A-7=autoinjector 7 doses; M-7 = manual injection 7 doses; Mean + SE (n = 7) one way ANOVA followed by Dunnett's comparison test with control group was carried out.

4. DISCUSSION

Several field equipments are available for the rapid detection of chemical agents and protection can be taken immediately. Additionally, medical protection can be taken using the first aid kit⁴. For the detection of the bio-threat

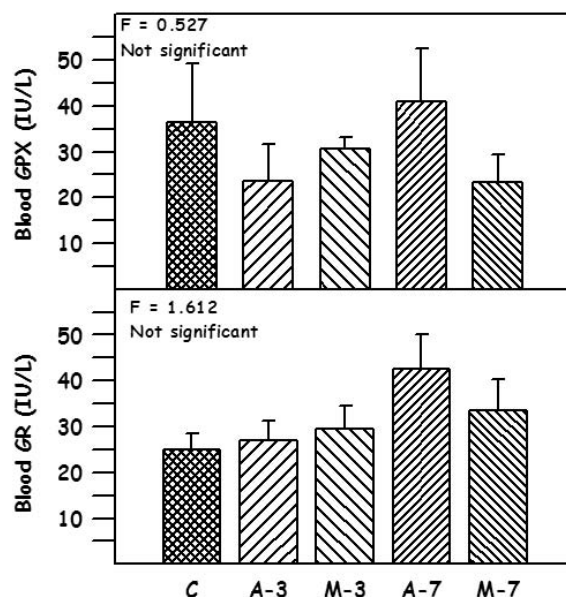


Figure 3. Effect of amikacin administered through autoinjector and manual injection by i.p. in rat on superoxide dismutase (SOD) and catalase (CAT), C = Control Group; A-3 = autoinjector 3 doses; M-3 = manual injection 3 doses; A-7 = autoinjector 7 doses; M-7 = manual injection 7 doses; Mean + SE (n = 7), one way ANOVA followed by Dunnett's comparison test with control group was carried out.

Table 1 . *In vitro* antibacterial activity of antibiotics individually and in combination against *Pseudomonas aeruginosa* and *Staphylococcus aureus*

Organism	Antibiotic	Zone of inhibition (mm) Mean ± SE (n=3)
<i>Pseudomonas aeruginosa</i> (Gram negative)	Amikacin	22.14 ± 0.48
	Cefazolin	19.55 ± 0.78
	Vancomycin	20.76 ± 1.05
	Amikacin + Cefazolin	22.25 ± 1.21
	Amikacin + Vancomycin	23.35 ± 0.69
	F = 2.80 P = 0.085	
<i>Staphylococcus aureus</i> (Gram positive)	Amikacin	18.25 ± 0.72
	Cefazolin	21.67 ± 0.69*
	Vancomycin	25.42 ± 0.46*
	Amikacin+Cefazolin	22.85 ± 0.90*
	Amikacin + Vancomycin	26.25 ± 0.95*
	F = 17.44 P < 0.001	

*Statistically significant from amikacin

Table 2 . Characteristics of bacterial bio threat organisms in categories A, B and C, and their drug of choice

Organism	Lethality	Human exposure	Grams reaction	Drugs -First choice	Drugs- other alternatives
Category A – Bacterial agents					
<i>Bacillus anthrax</i>	High	Aerosol	Gram positive	Ciprofloxacin, Doxycycline	Clindamycin, Rifampicin, Penicillin G
<i>Yersnia pestis</i>	High	Aerosol	Gram negative	Streptomycin, Doxycycline, Amikacin ¹⁷	Ciprofloxacin, Co trimaxazole, Gentamycin, Chloramphenicol
<i>Francisella tularensis</i>	Moderate	Aerosol	Gram negative	Gentamycin, Chloramphenicol, Amikacin ¹⁸	Ciprofloxacin, Doxycycline, Levofloxacin
Category B – Bacterial agents					
<i>Coxiella burnetti</i>	Low	Aerosol	Gram negative	Doxycycline	Tetracycline
<i>Brucella species</i>	Moderate		Gram negative	Doxycycline+ Streptomycin/ Gentamycin/ Amikacin ¹⁹	Doxycycline+ Rifampicin Cotrimoxazole+ Rifampicin Ciprofloxacin+ Rifampicin
<i>Burkholderia mallei</i>	Low	Aerosol	Gram negative	Cephalosporins Co trimoxazole Tetracycline	Aminoglycosides Amikacin ²⁰
<i>Rickettsia prowazekii</i>	Low		Gram negative	Doxycycline Tetracycline	Chloramphenicol Erythromycin
<i>Salmonella Species</i>	High to moderate	Food and water	Gram negative	Fluoroquinolones Chloramphenicol	Cefoperazone, Ceftriaxone Cotrimoxazole Amoxicillin Azithromycin
<i>Shigella dysenteriae</i>	Low	Food and water	Gram negative	Ciprofloxacin	Azithromycin, Ceftriaxone, Aminoglycosides Amikacin ²¹
<i>Escherichia coli</i>	Low	Food and water	Gram negative	Fluoroquinolones	Amoxycillin+ Clavulanic acid Amikacin ¹⁵ Cephalosporin, Tigecycline, Polymyxin B
<i>Vibrio cholerae</i>	High if untreated	Food and water	Gram negative	Doxycycline, Tetracycline	Cotrimoxazole, Chloramphenicol Fluoroquinolone Azithromycin, Amikacin ²²
Category C – Bacterial agents					
<i>Mycobacterium tuberculosis</i> MDR strain	High	Aerosol	Acid fast bacterium	Rifampicin Isoniazid Streptomycin	Quinolones Amikacin ²³

agents, field equipments are available but, their specificity is always questionable¹³. In such a situation, a prophylactic drug is essential and amikacin can be administered even in doubt. Table 2 lists the bio-threat agents as per Centers for Disease Control and Prevention (CDC) classification¹⁴. Category A includes bacterial agents that possess high risk to the public and national security. They can be easily spread or transmitted from person to person and result in high death rates and can cause public panic. Category B includes agents that are of second priority. They are easy to spread and produce moderate illness and low lethality. Whereas, category C includes agents which are emerging pathogens that could be engineered for mass spread in the future. They may cause high morbidity

and mortality rates and major health impact. Table 3 shows the bacterial agents, which are associated with field infections viz., military operations, terrorism, accidents and natural disasters¹⁴.

The drug of choice for each agent is also given in Tables 2 and 3. For the majority of the agents, particularly gram negative organisms, if a single antibacterial drug is required the preference is for amikacin. Hence, amikacin drug cartridge was developed for the existing autoinjector. This can be supplemented with oral doxycycline available in the first aid kit. In the present study, amikacin was tested against common field bacteria (gram negative and gram positive), *in vitro* for its effectiveness. Amikacin is an amino glycoside antibiotic derived

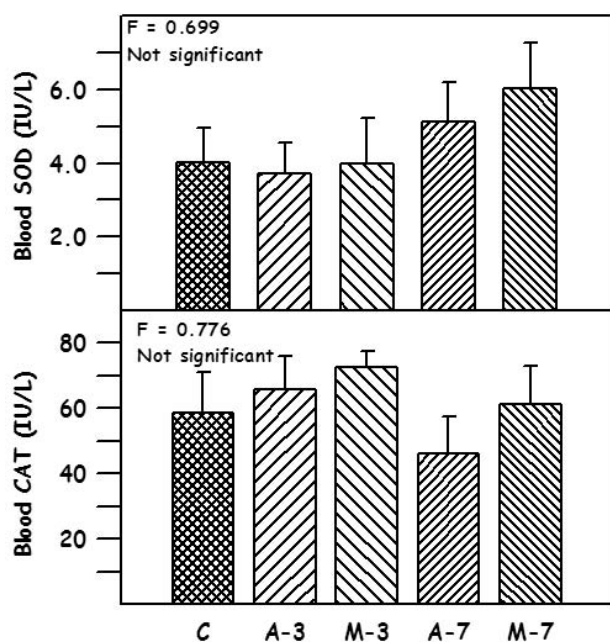


Figure 4. Effect of amikacin administered through autoinjector and manual injection by i.p. in rat on glutathione peroxidase (GPX) and glutathione reductase (GR), C = Control Group; A-3 = autoinjector 3 doses; M-3 = manual injection 3 doses; A-7 = autoinjector 7 doses; M-7 = manual injection 7 doses; Mean + SE (n = 7), one way ANOVA followed by Dunnett’s comparison test with control group was carried out.

from kanamycin and is highly effective against gram negative organisms as well as few gram positive organisms^{15,16}. It is also effective against gentamycin resistant strains. Although, amikacin is a very effective aminoglycoside against most of the gram negative organisms, it cannot be recommended for all the gram positive organisms. So a combination of antibiotics that can act against various gram negative and gram positive organisms will be useful in field administration. Combined antibiotic therapy is also useful in preventing resistance development and to enhance antibiotic efficacy¹⁵.

Unlike the chemical agents, for a specific detection the bio-threat agents require more time. Till the specific detection, prophylactic drug treatment is required. For gram negative organisms and also as a supportive treatment for gram positive organisms, amikacin is preferable. For viral agents and also for toxins, amikacin can be administered as a supportive therapy. The advantage of amikacin is its safety, stability and can be supplemented with other antibiotics when medical attention arrives. With the use of dual dose autoinjector, amikacin can be given to children and also for pet and farm animals. For biodefence, an antibiotic in an autoinjector is very essential. Amikacin is effective against most of the gram negative organisms. The present *in vitro* study revealed that a combination with either ceftazolin or vancomycin can be considered for future development as autoinjector cartridge for the existing autoinjectors.

The usability and tolerability of amikacin administered repeatedly through autoinjectors was earlier studied using

Table 3 . Characteristics of bacterial agents associated with field infections and their drug of choice

Organism	Lethality	Grams reaction	Drugs -first choice	Drugs- other alternatives
<i>Staphylococcus aureus</i>	Low	Gram positive	Penicillin G Methicillin Vancomycin Linezolid	Nafcillin Oxacillin Daptomycin Amikacin ²⁴
<i>Streptococcus pyogenes</i>	Low	Gram positive	Penicillin G or V	Cephalosporin Erythromycin Azithromycin Clindamycin
<i>Enterococcus faecalis</i>	Low	Gram positive	Penicillin G/ Vancomycin+ gentamycin	Ampicillin + Sulbactam + Imepenem
<i>Clostridium perfringens</i>	High	Gram positive [Anaerobe]	Metronidazole Gentamycin and amoxicillin	Clindamycin
<i>Clostridium tetani</i>	High	Gram positive [Anaerobe]	Penicillin G, Metronidazole	Clindamycin
<i>Pseudomonas aeruginosa</i>	Low	Gram negative	Ceftazidime Cefepime Amikacin ²⁴	Aztreonam Meropenem
<i>E. coli</i>	Low	Gram negative	Fluoroquinolones	Amoxycillin+ clavulanic acid Amikacin ¹⁵ Cephalosporin, Tigecycline, Polymyxin B
<i>Klebsiella</i> species	Low	Gram negative	Imipenem, Amikacin ¹⁵	Third generation cephalosporin, Cotrimoxazole
<i>Proteus</i> species	Low	Gram negative	Amikacin ¹⁵ Cotrimoxazole Imipenem	First generation cephalosporin

various haematological and biochemical parameters in animal models^{8,9}. The autoinjectors are self injectable medical devices and are intended for intramuscular or subcutaneous administration. To prove its safety, in the present study the amikacin autoinjector was administered intraperitoneally in rat, where the drug would be delivered with force on the abdominal vital organs. Hence, oxidative stress parameters were carried out in blood. The repeated administration of amikacin both by autoinjector and manual injection did not cause any change in the oxidative stress parameters. Though, autoinjector delivers the drug with a spray effect and with high force, no abnormality was detected in the rats when administered intraperitoneally, showing that autoinjectors are safe.

Conflict of Interest: None

REFERENCES

1. Porche, D.J. Biological and chemical bioterrorism agents. *J. Assoc. Nurs. AIDS. Care*, 2002, **13**(5), 57 - 64. doi: 10.1177/105532902236783
2. Ganesan, K.; Raza, S.K. & Vijayaraghavan, R. Chemical warfare agents. *J. Pharm. Bioallied Sci.*, 2010, **2**(3), 166 – 178. doi: 10.4103/0975-7406.68498
3. Thavaselvam, D. & Vijayaraghavan, R. Biological warfare agents. *J. Pharm. Bioallied. Sci.*, 2010, **2**(3), 179 - 188. doi: 10.4103/0975-7406.68499
4. Vijayaraghavan, R.; Bhaskar, A.S.B.; Gautam, A.; Gopalan, N.; Singh, A.K.; Beer Singh & Flora, S.J.S. A convenient first aid kit for chemical and biological agents and radiation exposure. *J. Environ. Biol.*, 2012, **33**(3), 673 - 681.
5. Friedl, K.E.; Hannan, C.J. Jr.; Schadler, P.W. & Jacob, W.H. Atropine absorption after intramuscular administration with 2-pralidoxime chloride by two automatic injector devices. *J. Pharm. Sci.*, 1989, **78**(9), 728 - 731. doi: 10.1002/jps.2600780905
6. Vijayaraghavan, R. Autoinjector device for rapid administration of life saving drugs in emergency. *Def. Sci. J.*, 2012, **62**(5), 307 – 31. doi: 10.14429/dsj.62.2317
7. Sheela, D.; Geetha, R.V.; Krishna Mohan, S. & Vijayaraghavan, R. A concept on the development of buprenorphine autoinjector for self and emergency administration. *Int. J. Pharm. Pharmaceut. Sci.*, 2015, **7**(8), 253 - 257.
8. Vijayaraghavan, R.; Selvaraj, R.; Krishna Mohan, S.; Gopi, P.G. & Tharani, C.B. Haematological and biochemical changes in response to stress induced by the administration of amikacin injection by autoinjector in animals. *Def. Sci. J.*, 2014, **64**(2), 99 - 105. doi: 10.14429/dsj.64.5032
9. Anitha Roy; Vijayaraghavan, R.; Geetha, R.V.; Anitha Magesh; Vishnu Priya, S.; Anusha, R.; Vidyalakshimi, U.; Raaga Namrata, K.; Krishna Mohan, S. & Madhan Chakkaravarthy, v. A comparative study of the effect of amikacin administered through autoinjector and manual injection on biochemical parameters in rats. *J. App. Pharm. Sci.*, 2016, **6**(2), 109 - 114. doi: 10.7324/JAPS.2016.60216
10. MacDougall, C. & Chambers, H.F. Aminoglycosides. *In Goodman and Gilman's the pharmacological basis of therapeutics*, Brunton, L.I.; Chabner, B.A & Knollmann, B.C. Ed. 12, Chapter 54. 2011, Mac Graw Hill, New York.
11. Villanova, P.A. National Committee for Clinical Laboratory Standards. Performance standards for antimicrobial disk susceptibility tests. Ed. 6. Approved Standard M2-A6. NCCLS, 1997.
12. Collee, J.G.; Miles, R.S. & Watt, B. Laboratory control of antimicrobial therapy. *In Collee, J.G.; Fraser, A.G.; Marmion, B.P. & Simmons, A (Eds): Mackie and McCartney Practical Medical Microbiology*, Ed. 14. Churchill Livingstone, New York, 1996, 151 - 178.
13. Hybl, J.D.; Tysk, S.M.; Berry, S.R. & Jordan, M.P. Laser-induced fluorescence-cued, laser-induced breakdown spectroscopy biological-agent detection. *Appl. Opt.*, 2006, **45**(34), 8806 - 8814. doi: 10.1364/AO.45.008806
14. Jernigan, D.B.; Raghunathan, P.L.; Bell, B.P.; Brechner, R.; Bresnitz, E.A.; Butler, J.C.; Cetron, M.; Cohen, M.; Doyle, T.; Fischer, M.; Greene, C.; Griffith, K.S.; Guarner, J.; Hadler, J.L.; Hayslett, J.A.; Meyer, R.; Petersen, L.R.; Phillips, M.; Pinner, R.; Popovic, T.; Quinn, C.P.; Reefhuis, J.; Reissman, D.; Rosenstein, N.; Schuchat, A.; Shieh, W.J.; Siegal, L.; Swerdlow, D.L.; Tenover, F.C.; Traeger, M.; Ward, J.W.; Weisfuse, I.; Wiersma, S.; Yeskey, K.; Zaki, S.; Ashford, D.A.; Perkins, B.A.; Ostroff, S.; Hughes, J.; Fleming, D.; Koplan J.P. & Gerberding, J.L. Investigation of bioterrorism-related anthrax, United States, 2001, Epidemiologic findings. *Emerg. Infect. Dis.*, 2002, **8**(10), 1019 – 1028. doi: 10.3201/eid0810.020353
15. Satoskar, R.S.; Rege, N.N. & Bhandarkar, S.D. Pharmacology and pharmacotherapeutics. Ed. 23. Popular Prakashan, New Delhi, 2013, 718 - 719.
16. Cunha, B.A. Aminoglycosides: Current role in antimicrobial therapy. *Pharmacotherapy*, 1988, **8**(6), 334-350. doi: 10.1002/j.1875-9114.1988.tb04092.x
17. Shcherbaniuk, A.I.; Makarovskaia, L.N.; Bugaeva, O.K. & Kasatkina, I.V. Antibiotics of the aminoglycoside group (gentamycin, sisomicin and amikacin) in the prevention and treatment of experimental plague. *Antibiot. Khimioter.*, 1992, **37**(5), 30 - 31.
18. Tynkevich, N.K.; Pavlovich, N.V. & Ryzhko, I.V. Comparative study of the effectiveness of amikacin and streptomycin in experimental tularaemia. *Antibiot. Khimioter.*, 1990, **35**(8), 35 - 37.
19. Ranjbar, M.; Keramat, F.; Mamani, M.; Kia, A.R.; Khalilian, F.O.; Hashemi, S.H. & Nojomi, M. Comparison between doxycycline-rifampin-amikacin and doxycycline-rifampin regimens in the treatment of brucellosis. *Int. J. Infect. Dis.*, 2007, **11**(2), 152 - 156. doi:10.1016/j.ijid.2005.11.007
20. Lipsitz, R.; Garges, S.; Aurigemma, R.; Baccam, P.; Blaney, D.D.; Cheng, A.C.; Currie, B.J.; Dance, D.; Gee, J.E.; Larsen, J.; Limmathurotsakul, D.; Morrow, M.G.; Norton, R.; O'Mara, E.; Peacock, S.J.; Pesik, N.; Rogers, L.P.; Schweizer, H.P.; Steinmetz, I.; Tan, G.; Tan, P.; Wiersinga, W.J.; Wuthiekanun, V. & Smith, T.L.

Workshop on treatment of and postexposure prophylaxis for *Burkholderia pseudomallei* and *B. mallei* Infection, 2010. *Emerg. Infect. Dis.*, 2012, **18**(12), e2.

doi: 10.3201/eid1812.120638

21. Wasfy, M.O. Isolation and antibiotic susceptibility of salmonella, Shigella and campylobacter from acute enteric infections in Egypt. *J. Hlth. Popul. Nutr.*, 2000, **18**(1), 33 - 38.
22. Mala, E.; Oberoi, A. & Alexander, V.S. Vibrio isolates from cases of acute diarrhoea and their antimicrobial susceptibility pattern in a tertiary care hospital. *Int. J. Basic. Appl. Sci.*, 2014, **3**(1), 35 - 37.
doi: 10.14419/ijbas.v3i1.1735
23. Gonzalo, X.; Casali, N.; Broda, A.; Pardieu, C. & Drobniowski, F. Combination of amikacin and doxycycline against multi drug – resistant and extensively drug – resistant tuberculosis. *Int. J. Antimicrobial agents.*, 2015, **45**(4), 406 - 412. doi: 10.1016/j.ijantimicag.2014.11.017
24. Nagoba, B.S.; Gandhi, R.C.; Hartalkar, A.R.; Wadher, B.J. & Selkar, S.P. Simple, effective and affordable approach for the treatment of burns infections. *Burns*, 2010, **36**(8), 1242 - 1247. doi: 10.1016/j.burns.2010.05.011

ACKNOWLEDGEMENTS

The author's are grateful to Director, DRDE (Gwalior) for all the support and M/s Neon Laboratories (Mumbai) for providing amikacin drug cartridges.

CONTRIBUTORS

Mrs R.V Geetha completed her MSc (Medical Microbiology) from Dr ALM Post Graduate Institute of Basic Medical Sciences, University of Madras. She is pursuing for her PhD at Saveetha University. Presently Working as Reader in the Department of Microbiology, Saveetha Dental College and Hospital. She has published 30 papers in the journals.

Mrs Anitha Roy has completed her MPharm from Dr Hari Singh Gour Vishwavidyalaya, Sagar. She is pursuing for her PhD at Saveetha University. She is presently Reader in the Department of Pharmacology at Saveetha Dental College and Hospital. She has published about 25 papers in the journals.

Dr S. Senthilkumar obtained his PhD (Biochemistry) from University of Madras and pursued postdoctoral research training in Ernst-Moritz-Arndt-University, Greifswald, Germany followed by Iowa State University and Stanford University, USA respectively. He is currently working as Assistant Professor in the Department of Research and Development, Saveetha University. He has published about 20 research papers in the journals.

Dr A.S.B. Bhaskar obtained his MSc and PhD in Biochemistry from Jiwaji University, Gwalior. Presently working as Scientist 'F' in Pharmacology and Toxicology Division, DRDE, Gwalior. He has 39 publications in national and international journals, one design patent, and one copyright. He has 24 years of research experience in diverse areas of research that includes natural toxins, chemical warfare agents, flow cytometry, molecular mechanisms in toxicology. He contributed in design and development of Autoinjectors and First Aid Kit for NBC emergency therapeutic use.

Dr R. Vijayaraghavan obtained his MSc and PhD in Medical Pharmacology. Presently he is working as Director-Research at Saveetha University, Chennai. He has about 250 research publications in reputed journals and about 60 patents, copyrights and designs. He has received *DRDO Agni Award of Excellence in Self Reliance - 2004* and *DRDO Titanium Trophy - 2007*. He has developed several products, viz., Personal decontamination kit, reusable autoinjectors, first aid kit for CBW agents, and insect repellents.