REVIEW ARTICLE

Human Pluripotent Stem Cells and Drug Discovery: A New Beginning

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ABSTRACT

Human pluripotent stem cells (hPSCs) offer unique opportunities to discover and develop a new generation of drugs. Their ability to differentiate into virtually any cell type renders them a cost-effective, renewable source of tissue-specific cell types capable of predicting human responses towards novel chemical entities. Using these improved *in vitro* models based on physiologically relevant human cell types could result in identifying highly precise and safe compounds, thereby reducing drug attrition rates. Moreover, ability to develop humanised disease models for patient-stratified drug screening makes hPSCs an impeccable tool in translational medicine. In this mini-review we focus on the positives and negatives of utilising hPSC-derived cell types as drug discovery platforms with special emphasis on cardio-, hepato- and embryotoxicity.

Keywords: Human pluripotent stem cells, cardiomyocytes, hepatocytes, drug discovery, cardiotoxicity, hepatotoxicity, disease modelling, embryotoxicity

1. INTRODUCTION

gradual Drug attrition; the reduction of pharmacological compounds while progressing along their development pipeline is the most pressing issue in the pharmaceutical industry over the last few decades. The main reason for drug attrition is the inappropriateness of a product with regard to either efficacy or safety. However, by the time these concerns are identified; new compounds have already entered clinical trials. In 2001, lack of efficacy and safety resulted in the withdrawal of $\sim 60\%$ of compounds which entered clinical trials¹. In 2004, the developmental cost of a marketable product was estimated at an escalating US\$ 900 million² and at this time, the US FDA had approved 36 new compounds which were later reduced³ to 20 by 2005. In order to reduce drug attrition rates as well as incurring developmental costs, evoking an in vitro humanised predictive system to evaluate chemical entities along their development pipeline is highly desirable.

Animal *in vitro* and *in vivo* test systems are currently being used to evaluate the efficacy and safety of a potential compound^{4,5}. However, due to species variation and pharmaco-toxicological differences, animal toxicological test systems are challenging, as results obtained from such models are not ideal representations of what may incur in humans⁶⁻⁸. The large number of animals required for comprehensive toxicological testing also poses a significant issue. The requirement for eliminating species variation and the constant pressure to minimise the use of test animals further emphasises the need to develop novel, robust and economical *in vitro* humanised models. This is not to say that human test systems are non-existent. However, human test systems which include immortalised cell lines and primary cell types, do not represent cell types in their natural state with primary cells losing tissue specific functions over time.

The derivation of human embryonic stem cells (hESCs) in the late nineties⁹ and induced pluripotent stem cells (hiPSCs) in 2007-08^{10,11} ushered in a new era in basic, translational and clinical research. The ability of human pluripotent stem cells (hPSCs) to self-renew and differentiate into virtually any cell type of the three germ layers provides an unparalleled advantage over somatic cell test systems^{12,13}. Their unlimited supply and reproducible differentiation¹⁴⁻¹⁷ into desired cell types allows for unique opportunities in drug development^{5,18-21}. In this mini-review, we direct our focus on the advantages and disadvantages of utilising hPSC-derived cell types as drug discovery platforms with special emphasis on cardio-and hepatotoxicity; forerunners for drug attrition as well as embryotoxicity.

2. HUMAN PSC-DERIVED CARDIOMYOCYTES

Cardiomyocytes are one of the most fundamental cell types in cardiac research, however severe limitations in obtaining workable human cardiomyocytes significantly hampers their use in pre-clinical drug discovery²². Moreover, due to their limited proliferation capacity, mature cardiomyocytes are incapable of undergoing longterm culture, requiring the need for repeated isolation of primary tissue necessary for in vitro functional analysis. Currently, canine cardiomyocytes are the most widely used pre-clinical model for assessing cardiotoxicity²³. Apart from being non-human, ethical concerns surrounding the killing of hundreds of dogs annually together with substantial costs are major limitations associated with this test system. Heterologous over-expression systems which include cell lines ectopically expressing specific cardiac ion channels are also utilised as in vitro test systems. However, over-expression of a single ion channel in isolation from a complex ion channel network within cardiomyocytes may prove to be disadvantageous. The market withdrawal of Vioxx further emphasises as to why in vitro humanised models which can predict efficacy and safety with high precision are mandatory^{5,23}.

Blockage of the potassium channel hERG which regulates the I_{κ} current is a key criterion for drug attrition. Proper regulation of $I_{\kappa r}$ is vital for normal cardiac repolarisation and prevention of QT prolongation. The QT interval represents the duration of ventricular depolarisation and subsequent repolarisation and prolongation of this interval to more than 450ms could trigger life-threatening arrhythmias²³. Pharmacological compounds such as thioridazine, nilotinib and arsenic trioxide are reported to cause QT prolongation. The FDA has thus recognised QT prolongation as an important variable which needs to be addressed during drug development while the International Conference of Harmonisation (ICH) Expert Working Group (EWG) for drug development has placed risk assessments for delayed ventricular repolarisation as standard procedure of preclinical evaluation of new chemical entities²⁴.

Cardiomyocytes differentiated from hPSCs open a new paradigm in regenerative medicine^{25,26}, disease modelling^{20,27}, drug discovery^{18-20,28} and understanding developmental phenomena^{17,29}. Similar to their adult counterparts, hPSC-derived cardiomyocytes express cardiac genes, structural proteins and ion channels^{17,30} as well as exhibit a functional excitation-contraction coupling system which could be modulated through $\beta 1/\beta 2$ adrenergic and muscarinic receptor agonists³¹. Evaluation of electrophysiological properties could be performed on single cells (patch clamp) or clusters (multi-electrode analysis; MEA) which allow for studying action potential (AP) and field potential duration (FPD) respectively (apart from other parameters). While the ability to assess the effect of a compound on a single ion channel (K⁺, Na⁺, Ca²⁺) or complex channel network (AP, FPD) enhances the prediction value of hPSC-cardiomyocytes, the absence of species variation renders these cells an impeccable humanised in vitro test system. A number of studies have demonstrated the potential application of hPSCcardiomyocytes in evaluating pharmacological safety^{23,32-} 36 . For example, while E-4031 (I $_{\rm kr}$ blocker) and Satalol (class III anti-arrhythmic agent; I $_{\rm Ks}$ blocker) were reported to induce QT prolongation, administration of quinidine

(class I anti-arrhythmic agent; I_{Na} blocker) and TTX (Na⁺ channel blocker) resulted in a dose-dependent reduction in conduction velocity^{18,32}.

With the establishment of hiPSC technology, patientspecific cardiomyocytes could be generated, providing unprecedented insight into the molecular mechanisms surrounding disease pathogenesis²⁰. Testing of compounds on patient-specific cardiomyocytes would result in a more accurate prediction with regard to efficacy and safety. Currently hiPSC models bearing various hERG mutations have been generated for studying congenital Long OT syndrome 2; a familial arrhythmogenic syndrome characterised by delayed repolarisation which could result inlifethreateningpolymorphicventriculartachycardia^{20,37,38}. While LQTS2 cardiomyocytes derived from patient-specific iPSCs recapitulate the disease phenotype (prolonged QT interval), treatment with ion channel modulators and β-blockers resulted in QT shortening and reduced the incidence of early after depolarisation (EAD) events. Targeting of the proteasomal complex has also proven to be successful in reversing the Long QT phenotype, showcasing the tremendous potential of hiPSCs in drug discovery²⁰.

Drawback surrounding hPSC-cardiomyocytes is their failure to completely develop a mature phenotype. Irregular sarcomeric organisation^{18,31}, multi-angular morphologies (spindle, round, triangular), absence of a T-tubule system³¹ and the expression of foetal proteins (e.g. SMA, Cx45)^{15,39,40} suggest hPSC-cardiomyocytes are structurally more foetal-like in nature. Spontaneous contractions, autoarrhythmic behaviour and smaller amplitudes of various currents indicate electrical immaturity as well. Based on these structural and electrophysiological disparities, when compared to adult cardiomyocytes, hPSC-cardiomyocytes seem to resemble a 16-week old foetal heart^{23,41} which may be disadvantageous during pharmacological evaluations. For example, dose-response studies have shown that hESC-cardiomyocytes respond to isoprenaline at ~10nM⁴², which is similar to a failing heart rather than a health one, which responds to \sim 1-2 nM depending on the age of the individual43.

When using hPSC-cardiomyocytes as a test system, evaluation of a compound focusses primarily on electrophysiological alterations (e.g. QT prolongation). It would be of utmost importance if hPSC-cardiomyocytes could also provide insight into compounds capable of inducing indirect cardiotoxicity. Treatment with anticancer drug trastuzumab (Herceptin); a monoclonal antibody administered for breast cancer resulted in a 4% increase in heart failure and decreased heart function by 10% especially in patients co-administered with anthracyclines⁴⁴. Since these side effects were unforeseen until phase III studies, adequate end-points should be developed to evaluate all-manner of cardiotoxicity (direct and indirect). To circumvent these issues, apart from electrophysiological alterations, the secretion of cardiac troponins and natriuretic peptides into culture medium have been used as indicators of potential cardiotoxicity^{45,46}. Improved understanding of candidate biomarkers would

therefore allow for heightened precision in predicting efficacy and safety of a novel chemical entity.

3. HUMAN PSC-DERIVED HEPATOCYTES

Targeting of liver metabolism, thereby modulating interplay between hepatocytes and other organs is considered a viable solution for metabolic ailments. However, more than 40% of new compounds fail their clinical phase trials due to unforeseen pharmacokinetic complications⁴⁷, rendering drug-induced liver toxicity a primary reason for attrition. It is estimated that 298 approved drugs cause hepatotoxicity as a side effect and while 6 compounds have been suspended or withdrawn, more than 250 compounds which are associated with acute liver failure still remain in the market⁴⁸. Hepatic death (apoptosis or necrosis) could be induced by a number of cellular factors (depletion of glutathione, binding to lipids and enzymes, direct targeting of mitochondrial complex and immune reactions)⁴⁹ with individual risk factors (genetic predisposition, differences in liver metabolism and environment) accounting for unforeseen complications⁵⁰.

Isolated human hepatocytes are considered suitable in vitro test systems for evaluating drug metabolism and toxicity, yet they are associated with severe limitations⁵¹. Apart from reduced proliferative capacity, primary hepatocytes retain physiological characteristics only for limited periods as most functions are lost during culture. Moreover, variation between donors (genetic polymorphisms, epigenetic status, health, diet and medications) makes unbiased evaluation and interpretation challenging. As with cardiomyocytes, species variation strongly hampers the utilisation of animal hepatocytes as test systems and while human hepatocellular carcinoma cell lines may be considered as alternatives to primary hepatocytes, these cell types exhibit partial hepatocyte characteristics^{47,52}. Humanised in vitro models capable of mirroring the complexity of hepatocytes with accurate prediction of candidate compounds is therefore highly desirable.

Human PSCs are an ideal source for obtaining functional hepatocytes and as reports suggest, more than 20 research groups are actively involved in attempts to optimize differentiation as well as maturation of hepatocytes. Combinations of growth factors and signal activators⁵³⁻⁶¹, co-culture techniques⁶²⁻⁶⁴ and various matrices⁶⁵⁻⁶⁷ have been implemented in attempts to induce hepatic differentiation. Once generated, hPSC-hepatocytes are reported to exhibit metabolic functionality such as glycogen storage, albumin secretion, urea synthesis, uptake and release of indocyanine green, AAT secretion and LDL uptake^{53,57,67-69}. Regardless of their metabolic functionalities, the most critical aspect for potential application of hPSChepatocytes in safety pharmacology and toxicology is their ability to metabolize drugs using phase I and II enzymes which facilitate drug metabolism⁵. Data based on hPSChepatocytes demonstrating phase I and II enzyme activity is still rudimentary with various discrepancies having been reported^{54,59-61,69-71}. The differentiation protocol

implemented, the hPSC line used, the quality of the standardised adult hepatocytes have been postulated as reasons for these reported discrepancies.

Despite many protocols being made available for the generation of hPSC-hepatocytes, down-stream cellular characterisation is based solely on gene and protein expression levels^{62,72}. The identity of hPSC-hepatocytes therefore remains an issue as non-hepatic cell types are also derived during the differentiation process. Development of a cost-effective, universally robust differentiation protocol would greatly assist in circumventing such issues. Furthermore, generation of hiPSCs from various ethnic groups and polymorphic variants as well as from patients with specific metabolic diseases would allow for population-based compound evaluation and influence the development of patient-stratified compounds.

4. HUMAN PSCS AND EMBRYOTOXICITY

Adverse effects caused by compounds on the reproductive system or developing embryo (growth retardation, malformations or death) is referred to as reproductive toxicology⁷³. While it would be ideal to model the reproductive cycle in vitro, its severe complexity renders it virtually impossible to do so. If the cycle were to be broken down into individual segments however, it may be possible to develop a suitable test system. Since this by itself is challenging, evaluation of embryotoxicity is still performed on pregnant animals including rats and rabbits. The large number of animals required for tests, including their offspring makes it a laborious and non-economical test system. As with cardio- and hepatotoxicity animal test systems, species variation would thwart accurate prediction of a given compound. An unfortunate example such as the thalidomide disaster has urged scientific communities to develop alternative methods for testing embryotoxicity, such as cell differentiation assays using either primary cells or immortalised cell lines^{73,74}.

Initially, mouse embryonic stem cells (mESCs) formed the basis of an innovative in vitro test system for screening potential embryotoxic compounds. Mouse ESCs could be differentiated into embryoid bodies (EBs) containing the three germ layers (ectoderm, mesoderm and endoderm), marking the onset of early mammalian embryogenesis, thereby establishing itself as a competent test system^{75,76}. When the mESC test (mEST) was compared against two other well-established in vitro models; rat whole-embryo culture (WEC) and the micromass (MM) test, in an international validation study funded by ECVAM (European Centre for the Validation of Alternative Methods), a 100% prediction rate in the evaluation of strong embryotoxic chemicals was achieved with all 3 in vitro test systems77. Acceptance of the mEST along with the WEC and MM test by the regulatory commission was therefore expected78; however, the Scientific Advisory Committee (ESAC) of the ECVAM felt that these *in vitro* test systems were not adequate⁷⁹. While several strategies have been implemented in attempts to improve mEST⁸⁰⁻⁹⁰, a major concern which remains is the species variation as it would be challenging to evaluate or interpret test results in a human context^{91,92}.

In an attempt to address species variation, hPSCs have been proposed as suitable models for investigating embryotoxicity^{91,93,94}. For instance, it was shown that while retinoic acid (RA) and 13-cis RA (13RA) exerted cytotoxic effects on hESCs, only the former compound induced cytotoxicity in mESCs despite both compounds being well-known agents of embryotoxicity⁹⁴. However, testing for cytotoxicity is not a sufficient end-point to establish reproductive toxicity and hence incorporation of biomarkers to monitor changes in early lineage transcripts and cardiac development has been proposed^{91,95}. In comparison to animal models, hPSCs have proven their effectiveness as an embryotoxic test system⁹⁶. For example, reversal of arsenic-induced embryotoxicity by monoisoamyl dimercaptosuccinic acid (MiADMSA) was not only observed in pregnant rats but in hESC-EBs as well. Furthermore, arsenic at non-cytotoxic concentrations of 0.1 ng/ml significantly down-regulated genes representing the germ layers which could be abrogated following MiADMSA treatment. These results suggest that while hPSC-EB test systems may be ideal for assessing embryotoxicity, its high level of sensitivity would assist in development of drugs which could revert or prevent the adverse effects of chemical entities.

While preliminary studies demonstrate an applicability of hPSCs as a viable embryotoxicity test system, certain issues are yet to be addressed. Human PSC lines have been reported to demonstrate lineage bias with propensities towards specific cell types during differentiation^{97,98}. Since lineage biases could lead to inaccurate predictions, it would be important to consider such intrinsic properties prior to selecting hPSC lines for compound evaluation studies. Ideally, hESC lines which demonstrate equal bias towards all three germ layers would be the most suitable candidates for such studies. When considering economical costs, commercial availability of high-throughput screening tools like AggreWells[™] (StemCell Technologies, CA) capable of generating large numbers of uniformed sised EBs could accentuate the screening process by allowing for evaluation of a large number of chemical entities. Finally, if hESC-based in vitro test systems could provide positive/ encouraging evaluations on drugs found to be embryotoxic following approval, then perhaps further developments in the field may be stimulated.

5. CONCLUSIONS

While hPSC implementation in therapeutic applications (regenerative medicine and disease modeling) has been extensively discussed over the past decade, only recently their potential in drug discovery is being realistically unravelled. The human origin, unlimited proliferative capacity and differentiation potential of hPSCs offer many advantages in biomedical research. Results obtained from proof-of-concept compound evaluation studies renders hPSCs a valuable tool for predicting efficacy and safety with high precision. For hPSCs to be successfully implemented as a robust drug discovery platform, issues needing to be addressed are (a) cost-effective and reproducible ways to generate purified cell populations for high-throughput screening, (b) development of universal and standardised assays for accurately evaluating toxicity of a chemical entity and (c) forging consensus between the scientific community, industry and regulators in developing guidelines for such evaluations.

Conflict of Interest : None

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